

## The essence of the TST procedure

1. Getting a fully “processable” tissue sample
2. Officially identifying the animal

It is important to understand that collecting a tissue sample is not just placing a tag. It is also delivering a processable tissue to the lab.

## Some important aspects are different from a standard tagging procedure:

- Good animal retention is essential
- Always use the tissue tag first (less animal resistance at sampling)
- Good positioning

## Correct use of the applicator is essential

Using the applicator as it is intended will help in correctly placing the tag and getting the best possible tissue for testing purposes.

## Clean and free from obstruction

Please keep the applicator clean and moving parts free from obstruction.

The light use of aerosol oil, especially before storage, is to be advised.

Please disinfect as required and in conjunction with good tagging practice.

## Do not separate the parts

The tags are supplied on plastic blisters – please do not separate the tag parts before the animal for which the tag is intended is retained and ready to receive the tag.

Every tube is marked with a unique bar code and the same unique number as the male and female tag parts. This permits secure registration and tracking on an individual level.

*Do not mix the parts up.*

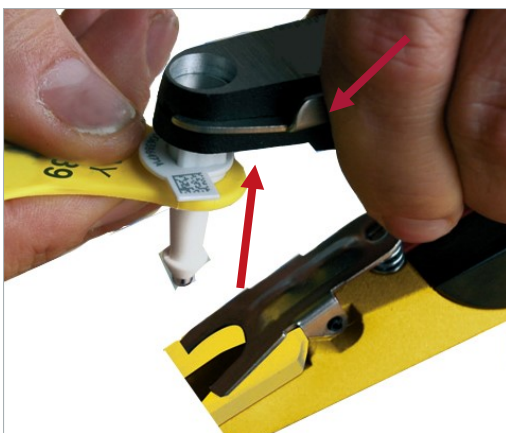


*The male and the female tag parts, as well as the needle and sample tube, are printed with the same number. This permits secure registration and tracking.*



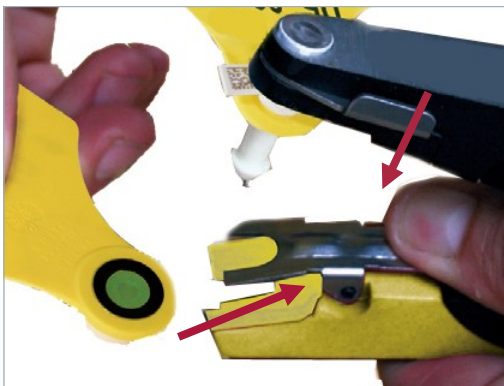
## 1. Inserting the male part into the pliers

Push on the safety button on the side of the jaws to enable the placement of the male part. Release the button to secure the male part in the jaw of the pliers.



## 2. Placing the female part into the pliers

Push the alloy pedal down to enable the placement of the female part. Release the pedal to clamp the female tag in place.



## 3. Applying the ear tag

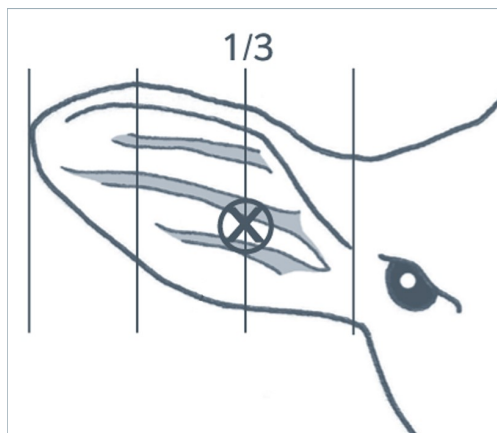
When applying the ear tags, always place the female part on the inside and the male part on the exterior side of the ear.

Make sure the animal is unable to move during the sampling.

When closing the pliers, use a firm and constant pressure.

You will hear two “clicks”. Allow the pliers to open immediately after the second “click” is heard.

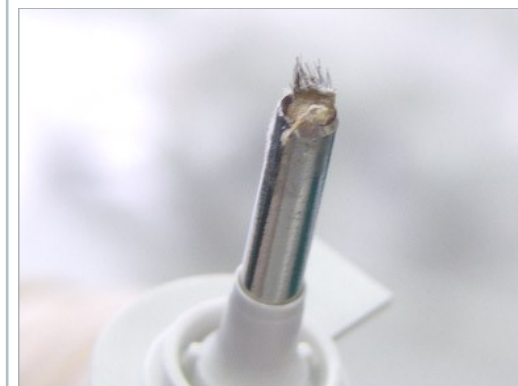
The ear tag will now be firmly attached to the ear.



## 4. Visible sample

The tag stays in the ear and the needle with tissue sample is retained in the applicator jaw after tagging is completed.

The tissue sample is clearly visible inside the needle.



If the pink cross shaped piston is still visible after sampling, the needle is empty.



If the pink piston is no longer visible, a tissue sample is successfully taken.





## 5. Applying the plastic tube onto the needle

It is good practice to hold the applicator in such a way that the needle points downwards while applying the sample tube. This to avoid the loss of the conservation fluid inside at the point of disrupting the water-proof seal.

Slide the tube onto the stainless steel needle while the needle is still inside the pliers. Push the tube firmly onto the needle.



Attach the two parts together by applying sufficient pressure until the plastic tube “clicks” into place on the base of the stainless steel tube part. Make sure both pieces (needle and tube) are inseparably fixed to each other.



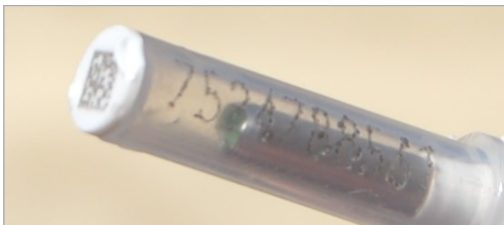
## 6. Removing the tube-needle unit from the pliers

Check that the needle is firmly sealed in the sample tube. Push on the safety button on the side of the jaws to enable the manual retraction of the tube-needle unit.



## 7. Tissue sample unit ready

The tube is clear and therefore it is possible to observe that a tissue sample has been successfully gathered into the tube. The tissue sample unit (plastic tube + stainless steel needle) is now ready for delivery at a selected laboratory.



## Sample storage

Collect the sealed tubes with enclosed needles in the collector as provided by your supplier or a suitable envelope, bag or container.

In preservation liquid, the samples can be preserved up to one month at room temperature, without any impact on both BVDV diagnoses as DNA extraction on genomic analysis.

The samples can also be stored in a refrigerator, at ca. 4 °C, up to one month.

## Packing and sending

Collect the tubes with enclosed needles in the collector as provided by your supplier or a suitable envelope or container. A strong padded envelope is ideal.

Please notice that correct packaging depends on the local legislation for sending biological materials.

Different countries have different demands for such postal services.



## OS ID TST can be used for:

- BVDV diagnostics (preservation liquid not essential)
- Genomic index calculations
- Genomic trades
- Scrapie resistance

## Areas of use for the OS ID TST

- Breeding work and selection
- Research
- Disease prevention

## Animal welfare

The OS ID TST is gentle on animals. With a needle diameter of just 3 mm, only a small hole is left in the ear, through which the tag is securely attached.

## Animal restraint

Animal restraint is essential in taking a good tissue sample. Make sure the animal is unable to move during the sampling process/identification.

